

## Note

### Synthesis of a C-glycosyl amino acid analogue of *O*-( $\beta$ -D-xylopyranosyl)serine

Ladislav Petruš<sup>1</sup> and James N. BeMiller

*The Whistler Center for Carbohydrate Research, Purdue University, West Lafayette, Indiana 47907 (USA)*  
(Received August 9th, 1991; accepted December 12th, 1991)

C-Glycosyl analogues of naturally occurring compounds have been prepared for a variety of reasons, primarily as inhibitors of glycosidic bond hydrolases or to prepare a more stable form of an active principle. An interesting class of C-glycosyl compounds are the C-glycosyl amino acids. Previously prepared have been  $\alpha$ -C-glycosyl- $\alpha$ -amino acids<sup>1,2</sup>. Especially interesting would be those C-glycosyl amino acids that are analogues of points of saccharide attachment in glycoproteins, proteoglycans, and protein-polysaccharides<sup>3</sup>.

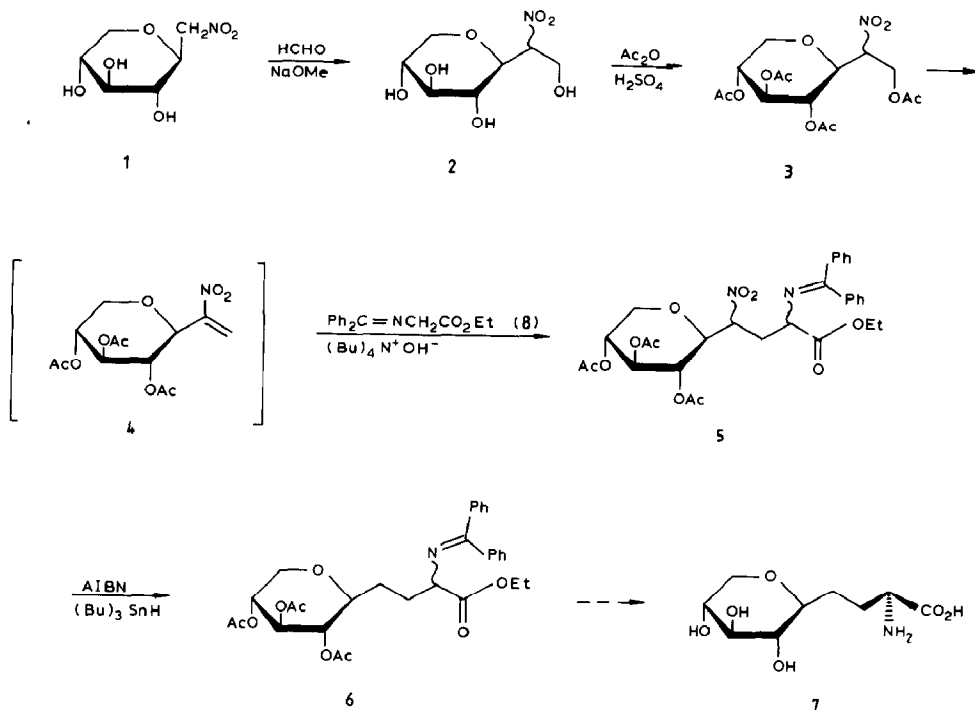
Herein, we describe the synthesis of intermediates related to one such compound, the C-glycosyl ( $-\text{CH}_2-$ ) analogue (**7**) of *O*-( $\beta$ -D-xylopyranosyl)-L-serine. Synthesis began with ( $\beta$ -D-xylopyranosyl)nitromethane<sup>4</sup> (**1**) \*. (Glycopyranosyl)nitromethanes, also known as 2,6-anhydro-1-deoxy-1-nitroalditols, are convenient starting compounds for the preparation of C-glycosyl compounds via nitroaldol condensation with an appropriate aldehyde<sup>5</sup>, a reaction analogous to the classical nitromethane synthesis. Another approach, the addition of C-nucleophiles to  $\alpha,\beta$ -unsaturated nitro sugars, has been used to prepare branched-chain sugar derivatives from 2,3-dideoxy-3-nitro-olefins<sup>6,7</sup>. This communication extrapolates this approach to 2-glycopyranosyl-2-nitroethanols<sup>8</sup>, other potential precursors of nitro-olefins, and describes a simple approach to **7**.

Nitroaldol addition of **1** to formaldehyde provided a mixture of epimeric 2-nitro-2-( $\beta$ -D-xylopyranosyl)ethanols (**2**). Formation of their insoluble sodium salts was a decisive step in their synthesis, as normally all the acidic  $\alpha$ -protons of primary nitroalkanes react with formaldehyde<sup>9–14</sup>. In fact, **1** does undergo double electrophilic addition of formaldehyde in aqueous alkaline solution (unpublished result). Simultaneous use of carbon dioxide buffering and a strongly acidic cation-

*Correspondence to:* Professor J.N. BeMiller, The Whistler Center for Carbohydrate Research, Purdue University, West Lafayette, IN 47907 USA.

<sup>1</sup> Permanent address: Institute of Chemistry, Slovak Academy of Sciences, 84238 Bratislava, Czechoslovakia.

\* Compound **1** is properly named as 2,6-anhydro-1-deoxy-1-nitro-D-gulitol.



exchange resin produced **2** from its sodium nitronate form and prevented further condensation.

Per-*O*-acetylation produced no change in the ratio of epimers.

Olefin **4**, expected as a dehydroacetoxylation product of **3**, could not be isolated. When isolation was attempted, a new product was formed, probably in a reaction sequence beginning with the opening of the anhydro ring. Therefore, **3** was allowed to react with a glycine nucleophile generated from ethyl *N*-diphenylmethyleneglycinate<sup>15</sup>, but by a somewhat different procedure than that reported<sup>16</sup>. The coupling reaction afforded the expected product in 50% yield as a mixture of all four possible diastereomers. Crystallization gave one isomer (**5**), one of the principal ones. The configuration of this isomer at the CH–NO<sub>2</sub> and CH–N= centers is being determined by crystallographic analysis.

For denitration, a radical process employing tributyltin hydride and azobis(isobutyronitrile) (AIBN) was used<sup>17</sup>. Proof of the structure of **6** was given by mass and <sup>13</sup>C-NMR spectra. The overall yield of **6** (single isomer) from **1** was 4.2%.

## EXPERIMENTAL

*Addition of (β-D-xylopyranosyl)nitromethane (1) to formaldehyde.*—Formaldehyde (0.6 g, generated by thermal decomposition of paraformaldehyde) was intro-

duced to a solution of **1** (1.93 g, 10.0 mmol)<sup>2</sup> in Me<sub>2</sub>SO (10 mL). A solution of sodium methoxide (prepared from 0.35 g of Na) in MeOH (10 mL) was added. The mixture containing a precipitate was stirred (1 min). Then, 1-butanol (70 mL) was added. (This mixture could be kept several days at 5° without changes.) The precipitate was collected by filtration, washed with 1-butanol (5 × 10 mL), and added under vigorous stirring to a suspension of a strongly acidic cation-exchange resin (H<sup>+</sup> form, 50 mL, exchange capacity 1.5 mequiv/mL) and crushed solid CO<sub>2</sub> (100 g) in water (200 mL). Filtration and evaporation of the filtrate provided a syrup (2.14 g, 96%) containing **2** (epimeric ratio, 2:1 by <sup>13</sup>C-NMR spectroscopy) and starting material (~5%).

**Acetylation of 2.**—Compound **2** (syrupy mixture, 1.40 g, 6.28 mmol) was dissolved in MeOH (3 mL) and added dropwise into acetic anhydride (50 mL) containing concentrated H<sub>2</sub>SO<sub>4</sub> (4 drops) at 30–40° with stirring. After the final addition, the mixture was stirring an additional 2 h. The clear solution was then poured into ice and water (300 mL), and the mixture was stirred 3 h. Extraction of the water with CHCl<sub>3</sub>, drying (Na<sub>2</sub>SO<sub>4</sub>), and evaporation afforded tetraacetate **3** (1.69 g, 69%, epimeric ratio, 2:1) (identity confirmed by <sup>13</sup>C-NMR spectroscopy).

**Reaction of 3 and 8.**—A solution of *N*-diphenylmethyleneglycine ethyl ester (**8**) (0.53 g, 2.0 mmol)<sup>15</sup> and tetrabutylammonium bromide (0.64 g, 2.0 mmol) in CHCl<sub>3</sub> (50 mL) was shaken with aq M NaOH (50 mL) for 10 min. The CHCl<sub>3</sub> layer was removed and dried (Na<sub>2</sub>SO<sub>4</sub>), and to it was added **3** (0.78 g, 2.0 mmol). The mixture was stirred 16 h at room temperature. Column chromatography (Silica Gel-60, 4:3 v/v hexane–EtOAc) provided a fraction from which a crystalline diastereoisomer of ethyl 2-(diphenylmethyleamino)-4-nitro-4-(2,3,4-tri-*O*-acetyl-β-D-xylopyranosyl)butanoate (**5**, 0.16 g, 13%) was obtained: mp 149–151°; [α]<sub>D</sub><sup>25</sup> –103° (c 1.65, CHCl<sub>3</sub>); *m/z* 599 [M + H]<sup>+</sup> (FAB mass spectrum). <sup>13</sup>C-NMR (50.309 MHz, CDCl<sub>3</sub>, internal Me<sub>4</sub>Si), δ: 14.10 (ethyl CH<sub>3</sub>), 20.51, 20.67 (acetyl CH<sub>3</sub>), 29.58 (C-3), 60.96 (ethyl CH<sub>2</sub>), 61.38 (C-5'), 66.81 (C-2), 68.46 (C-2'), 69.15 (C-4'), 73.76 (C-3'), 78.89 (C-1'), 83.45 (C-4), 127.76, 128.08, 128.55, 128.82, 130.76, 135.56, 138.74 (aryl C), 169.16, 169.68, 170.27, 170.60 (C-1, acetyl C=O), 172.74 (C=N).

**Anal.** Calcd for C<sub>30</sub>H<sub>34</sub>N<sub>2</sub>O<sub>11</sub>: C, 60.19; H, 5.73; N, 4.68. Found: C, 60.34; H, 5.83; N, 4.52.

**Denitration of 5.**—A mixture of **5** (122 mg, 0.2 mmol), AIBN (33 mg, 0.2 mmol), tributyltin hydride (0.55 mL, 2 mmol), and toluene (0.3 mL) was stirred at 110° under an atmosphere of N<sub>2</sub> for 1 h<sup>17</sup>. Column chromatography (Silica Gel-60, 5:3 v/v hexane–EtOAc) afforded ethyl 2-(diphenylmethyleamino)-4-(2,3,4-tri-*O*-acetyl-β-D-xylopyranosyl)butanoate (**6**, 26 mg, 24%; overall yield from **1** 2.1%); [α]<sub>D</sub><sup>25</sup> –84.1° (c 1.35, CHCl<sub>3</sub>); *m/z* 554 [M + H]<sup>+</sup> (FAB mass spectrum). <sup>13</sup>C-NMR (50.309 MHz, CDCl<sub>3</sub>, internal Me<sub>4</sub>Si), δ: 14.21 (ethyl CH<sub>3</sub>), 20.56, 20.73 (acetyl CH<sub>3</sub>), 27.59 (C-4), 28.89 (C-3), 60.93 (ethyl CH<sub>2</sub>), 64.52 (C-1'), 66.65 (C-2), 69.36 (C-5'), 72.07 (C-4'), 73.82 (C-2'), 77.52 (C-3'), 128.03, 128.49, 128.80, 130.03, 130.40, 136.27, 139.37 (aryl C), 169.59, 169.82, 170.40, 170.53 (C-1, acetyl C=O),

171.95 (C=N). As the ( $\beta$ -D-xylopyranosyl)methyl group is identical to the 1,5-anhydro-6-deoxy-L-glucitol-6-yl group, assignment of the xylopyranosyl carbon atoms was made from data for 1,5-anhydro-D-glucitol and methyl 6-deoxy- $\alpha$ -D-glucopyranoside<sup>18</sup>.

#### ACKNOWLEDGEMENTS

The authors acknowledge the provision of travel funds by the National Academy of Sciences of the U.S.A. and the Czechoslovak Academy of Sciences that made it possible for L. Petruš to work in the Whistler Center for Carbohydrate Research. Mr. John A. Bohn is thanked for determining the NMR spectra.

#### REFERENCES

- 1 R.H. Hall, K. Bischofberger, S.J. Eitelman, and A. Jordaan, *J. Chem. Soc., Perkin Trans. 1*, (1977) 743–753.
- 2 G. Simchen and E. Pürkner, *Synthesis*, (1990) 525–527.
- 3 N. Sharon, *Complex Carbohydrates, Their Chemistry, Biosynthesis, and Functions*, Addison-Wesley, Reading, MA, 1975.
- 4 L. Petruš, S. Bystrický, T. Sticzay, and V. Bílik, *Chem. Zvesti*, 36 (1982) 103–110.
- 5 O.R. Martin and W. Lai, *J. Org. Chem.*, 55 (1990) 5188–5190.
- 6 H.H. Baer and K.S. Ong, *Can. J. Chem.*, 46 (1968) 2511–2517.
- 7 T. Sakakibara, S. Kumazawa, and T. Nakagawa, *Bull. Chem. Soc. Jpn.*, 43 (1970) 2655.
- 8 M. Petrušová, E. Lattová, M. Matulová, and L. Petruš, *Chem. Pap.*, 46 (1992) in press.
- 9 I.M. Gorski and S.P. Makarow, *Ber.*, 67 (1934) 996–1000.
- 10 H.B. Hass and B.M. Vanderbilt, U.S. Pat. 2 139 120 (1937); *Chem. Abstr.*, 33 (1939) 2149.
- 11 B.M. Vanderbilt and H.B. Hass, *Ind. Eng. Chem.*, 32 (1940) 34–38.
- 12 H.H. Baer and L. Urbas, in H. Feuer, (Ed.), *The Chemistry of the Nitro and Nitroso Groups*, Interscience, New York, 1970, pp. 75–200.
- 13 P.A. Wade and R.M. Giuliano, in H. Feuer and A.T. Nielsen, (Eds.), *Nitro Compounds*, VCH, New York, 1990, pp. 137–265.
- 14 R.F.B. Cox, U.S. Pat. 2 301 259 (1939); *Chem. Abstr.*, 37 (1943) 2017.
- 15 M.J. O'Donnell and R.L. Polt, *J. Org. Chem.*, 47 (1982) 2663–2666.
- 16 L. Ghosez, J.-P. Antoine, E. Deffense, M. Nivarro, V. Libert, M.J. O'Donnell, W.A. Bruder, K. Willey, and K. Wojciechowski, *Tetrahedron Lett.*, 23 (1982) 4255–4258.
- 17 N. Ono, H. Miyake, and A. Kaji, *J. Org. Chem.*, 49 (1984) 4997–4999.
- 18 K. Bock and C. Pedersen, *Adv. Carbohydr. Chem. Biochem.*, 41 (1983) 27–66.